



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 1

APPROVED

**SOURCE: USAHA/AAVLD COMMITTEE ON ANIMAL EMERGENCY
MANAGEMENT**

SUBJECT MATTER: Veterinary License Reciprocity in Emergencies

BACKGROUND INFORMATION:

Large-scale animal emergency disasters can occur during events such as hurricanes, floods, fires, and disease outbreaks. These events have often exhausted in-state resources requiring states to reach out to other states and national organizations to assist in response and recovery efforts. The veterinary community has organized itself sufficiently in recent years to respond to such requests for assistance. A limiting factor in fulfilling requests for assistance is the lack of a standardized means of addressing reciprocal licensure during emergencies. Inconsistencies in states' licensing board processes as well as refusal of some boards to recognize out-of-state licenses during emergencies has led to delays in providing assistance when critically needed.

Nationally, there are two professional and legal means for addressing this issue. First, the Emergency Management Assistance Compact (EMAC) is a congressionally ratified mutual aid compact that legally establishes a national system to facilitate the deployment of resources across state lines during an emergency or disaster. To date, all fifty states, the District of Columbia, Puerto Rico, Guam, and the United States Virgin Islands are EMAC members. EMAC is state law; therefore, in most cases, a licensing board does not supersede state law. The state emergency management agencies (EMAs) within the EMAC Member States are responsible for the implementation of EMAC. Second, request of licensed veterinary professionals via non-EMAC processes such as Memoranda of Agreement (MOA) between state emergency management and recognized entities or organizations allows for specific requirements for deployment to be outlined in advance which streamlines the license reciprocity processes. These means are both effective and protective due to the national veterinary licensure examination and continuing education requirements in place to ensure continuity and standardization of the practice of veterinary medicine in the United States. The American Veterinary Medical Association (AVMA) Model Veterinary Practice Act has a provision allowing for emergency licensing of out-of-state veterinarians. This language could be adapted for state use.

RESOLUTION:

The United States Animal Health Association urges the American Association of Veterinary State Boards to develop and distribute to veterinary state boards a position statement supporting processes that enable veterinary medical personnel to operate under reciprocal veterinary medical licensure when emergency assistance is requested by their state and is in accordance with state emergency management laws, regulations, and guidelines.

TO: State Boards of Veterinary Medicine

FROM: The American Association of Veterinary State Boards

RE: Veterinary License Reciprocity in Disasters

The American Association of Veterinary State Boards (AAVSB) has received a Resolution from the US Animal Health Association (USAHA) requesting us to issue a statement to our members in support of veterinary license reciprocity in disaster situations. Through USAHA, it has come to our attention that the process of enabling veterinarians to practice across state lines in response to a disaster is a concern for state veterinary licensing boards, veterinary responders, state emergency management officials, and state animal health officials. They shared that in recent disasters, inconsistencies in states' licensing board processes as well as refusal of some boards to recognize out-of-state licenses during emergencies have led to delays in providing critical assistance to animals and animal owners.

The USAHA is an organization of personnel from State and Federal governments, universities, industry, and other concerned groups that addresses issues of animal health and disease control, animal welfare, food safety and public health. It is a clearinghouse for new information and methods, which may be incorporated into laws, regulations, policy, and programs. It develops solutions of animal health-related issues based on science, new information and methods, public policy, risk/benefit analysis and the ability to develop a consensus for changing laws, regulations, policies, and programs. The mission of USAHA is to develop and promote sound animal health solutions for public good. Since 1897, USAHA science-based committees have deliberated each year regarding current issues to create "Resolutions" which are recommendations aimed at solving critical animal and public health problems.

AAVSB received the Resolution about license reciprocity in disasters from the USAHA Committee on Animal Emergency Management (CAEM). It includes the language excerpted below where CAEM describes two "professional and legal means" which they believe support, along with national veterinary licensure examination and continuing education requirements in each state, the concept that effective protections are already in place to allow consideration for states to develop processes to allow temporary licensure for invited veterinary professionals.

First, the Emergency Management Assistance Compact (EMAC) is a congressionally ratified mutual aid compact that legally establishes a national system to facilitate the deployment of resources across state lines during an emergency or disaster. To date, all fifty states, the District of Columbia, Puerto Rico, Guam, and the U.S. Virgin Islands are EMAC members. EMAC is state law; therefore, in most cases, a licensing board does not supersede state law. The state emergency management agencies (EMAs) within the EMAC Member States are responsible for the implementation of EMAC. Second, request of licensed veterinary professionals via non-EMAC processes such as Memoranda of Agreement (MOA) between state emergency management and recognized entities or organizations allows for specific requirements for deployment to be outlined in advance which streamlines the license reciprocity processes."

It may also be noted that the AVMA Model Veterinary Practice Act has a provision allowing for emergency licensing of out of state veterinarians. The language is found in Section 6 paragraph 15:

15. A veterinarian licensed or a veterinary technician credentialed in another state may practice in the State during an emergency or natural disaster within the scope and location of assigned veterinary medical duties of the response efforts without written examination or other qualification if:
 1. an official declaration of the disaster or emergency has been made by the Governor or the delegated State official; and

2. an official invitation has been extended to the veterinarian or veterinary technician for a specified time by the authority that has jurisdiction for coordinating the animal/agricultural issues in the State during emergencies either within or outside the Emergency Management Assistance Compact (EMAC).

While the AAVSB is not in a position to comply with state law requirements, it is understood that many state veterinary boards may be compelled to do so. Therefore, we have decided to support the recommendation of the USAHA CAEM and we encourage state boards of veterinary medicine to identify stakeholder groups in your states to identify mutually-beneficial processes for enabling the practice of veterinary medical personnel by reciprocity as requested.

This encouragement is made with the understanding that veterinary professionals licensed in other states who are seeking temporary licensure to practice in a disaster-affected state have been invited by state animal officials to assist for disaster response purposes only and are working within the outline of the law and the state's veterinary practice act. The AAVSB does not support self-deployment of veterinarians into disaster situations.



RESOLUTION NUMBER: 2

APPROVED

SOURCE: USAHA/AAVLD COMMITTEE ON ANIMAL EMERGENCY MANAGEMENT

SUBJECT MATTER: Radiological Incident Response and Resources

BACKGROUND INFORMATION:

With more than 100 fixed nuclear facilities nationwide, states must be prepared to assist citizens in the event of a site emergency. Public health and other partners will look to animal/agricultural responders for resources needed for service animals and pets. State animal/agriculture emergency planners have identified a severe lack of these resources and therefore a serious gap in our national animal response capability.

Since October 2006, the Pet Evacuation and Transportation Standards (PETS) Act has required local and state emergency plans to include citizens with service animals and pets before, during, and after disasters of all types. Citizens evacuated during a radiation emergency event arriving at reception centers with their service animals and pets will require triage, radiation monitoring, external decontamination, and post-decontamination services and support. Trained personnel, standardized protocols and equipment (including personal protective equipment) must be in place to provide these services. Because only a very limited number of persons have received animal decontamination training at both state and federal levels, resources would be immediately overwhelmed in a disaster.

The United States Department of Health and Human Services and National Disaster Management System (HHS/NDMS) have proven experience at the development and maintenance of personnel resources such as the National Veterinary Response Team (NVRT) to assist states. We believe HHS/NDMS/NVRT provides the ideal solution to fill this critical response gap by development of the following resources: caches of equipment to include mobile animal decontamination portals; personnel teams with current training in animal decontamination techniques; and delivery of guidance and standardized training that can build local response capability to assist animal/agricultural and public health emergency responders and citizens at local, state and federal levels.

The Federal Emergency Management Agency (FEMA) Radiological Emergency Preparedness (REP) Program coordinates the national effort to provide state, local, and tribal governments with relevant and executable planning, training, and exercise guidance and policies necessary to ensure that adequate capabilities exist to prevent, protect against, mitigate the effects of, respond to, and recover from incidents involving

commercial nuclear power plants. Following a request from the American Veterinary Medical Association (AVMA) in 2014 to suggest that the REP Program utilize available pet decontamination guidelines to expand the REP program guidelines, the January 2016, REP Program Manual states “FEMA encourages offsite response organizations to plan for the reality that in an emergency, many evacuees will arrive at reception centers with their pets” and “no specific guidance on the radiological monitoring and decontamination of household pets currently exists.”

The United States Department of Homeland Security Science and Technology Directorate is capable of performing research that could produce scientific data that could be used to develop best practices for animal decontamination.

This Resolution was originally addressed to the Department of Health and Human Services in 2014, stating: “The United States Animal Health Association urges the Department of Health and Human Services to develop and maintain personnel, equipment, and training resources, especially those needed for pet and service animal decontamination, to supplement state animal response in radiation emergencies and all-hazards events.” The issue is as relevant in 2016 as it was in 2014.

RESOLUTION:

The United States Animal Health Association (USAHA) urges the United States Department of Homeland Security (DHS), Science and Technology Directorate, to develop and perform research to produce data related to effective methods of animal decontamination in radiological events. Furthermore, DHS is urged to coordinate with the Federal Emergency Management Agency (FEMA) Radiological Emergency Preparedness (REP) Program to apply this data toward development of best practices for decontamination of animals. Lastly, USAHA urges DHS and FEMA REP to partner with the Department of Health and Human Services National Disaster Management System/National Veterinary Response Team programs to develop and deliver training courses to fill the gaps in nuclear event response capabilities that currently exist in local and state jurisdictions.



RESOLUTION NUMBER: 3

APPROVED

SOURCE: USAHA/AAVLD COMMITTEE ON ANIMAL EMERGENCY MANAGEMENT

SUBJECT MATTER: Resource Typing for Animal Emergency Response

BACKGROUND INFORMATION:

The Post-Katrina Emergency Management Reform Act guided the Federal Emergency Management Agency (FEMA) to reach an “understanding with non-federal officials” on standards for credentialing of personnel and typing of response resources. Maintenance of the National Incident Management System (NIMS), as required under Department of Homeland Security (DHS) Presidential Directive 5 (HSPD-5), included establishment of the National Integration Center (NIC) which has the responsibility for standards and credentialing.

Beginning in 2007 and meeting regularly for more than five years, the FEMA Animal Emergency Response Working Group (AERWG) produced volumes of collaborative work products which included descriptions, specifications, and training requirements for at least 25 critical individual animal emergency response (AER) positions and several AER teams. The group included animal/agriculture emergency managers and responders with experience in disasters across the United States along with other national resource typing experts. The entire body of AERWG work, much of which had been vetted nationally, was never published.

Various groups, including state animal health officials, have worked independently to create AER resource typing guidelines for disaster events. The Southern Agriculture and Animal Disaster Response Alliance (SAADRA), a 13-state planning and coordination group formed in 2006, expanded the FEMA “508-1” list with detailed descriptions of 11-Type II through IV animal emergency response teams. Later, SAADRA and the National Animal Rescue and Sheltering Coalition (NARSC), an organized alliance of national animal responders, modified some of the team specifications. In 2014, the National Association of State Animal and Agriculture Emergency Programs (NASAAEP), a national group of animal and agriculture emergency managers appointed by chief state animal health officials in every state, amicably discussed a plan with the NIC Coordinator to begin a project of revising and accepting typing standards for these critical resources.

It is understood that a full inventory of AER resources will likely remain a living document requiring periodic revision. An example of this is the discovery of the need for a Case Manager position that surfaced in a recent animal disease event. We need to move forward to adopt resource typing guidelines to improve our national response capabilities.

RESOLUTION:

The United States Animal Health Association urges the Federal Emergency Management Agency (FEMA) National Integration Center (NIC) to do the following:

- Publish and announce a temporary endorsement of the 11- typed animal emergency response (AER) teams created by the Southern Agriculture and Animal Disaster Response Alliance and the National Animal Rescue and Sheltering Coalition in place of the currently published FEMA 508-1;
- Assemble a small team of AER subject matter experts, including former Animal Emergency Response Working Group (AERWG) members, to revise the AERWG draft products within a 6-month time frame; and
- Implement a system to allow revision of AER resources, as needed, every 3 years.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 4, 21 and 26 Combined

APPROVED

SOURCE:

**COMMITTEE ON ANIMAL EMERGENCY MANAGEMENT
COMMITTEE ON IMPORT, EXPORT AND
INTERNATIONAL STANDARDS
COMMITTEE ON BIOLOGICS & BIOTECHNOLOGY**

SUBJECT MATTER:

National Foot-and-Mouth Disease Preparedness

BACKGROUND INFORMATION:

Foot-and-Mouth Disease (FMD) is the most contagious and economically destructive disease of livestock. An FMD event in the United States will have severe, profound and long lasting negative impact on the United States agriculture and general economy. The United States Department of Agriculture (USDA) estimates that economic losses due to an FMD event in the United States will range from \$15 billion to \$100 billion per year (Source: USDA FMD Vaccination Policy in the United States, September 2014). Recent experiences in the United States with foreign animal disease outbreaks (porcine epidemic diarrhea virus (PEDv) and H5 type high pathology avian influenza (HPAI)) underscore the need for preparedness in dealing with high consequence animal disease impacting agriculture. In collaboration with animal agriculture stakeholders, allied industry, academia, State and other Federal agencies, the USDA continues to progress on FMD preparedness and response planning.

Previously applied FMD disease mitigation through culling-to-control methods are not considered effective and practical for the scale and advancement of the United States livestock industry. Emergency FMD vaccination control measures with effective elimination strategies are the most viable option for minimizing the economic impact of the disease. Should FMD become endemic after an outbreak in North America, control of the disease with vaccination will likely assure some level of continuity of business for United States livestock producers.

The September 2014 USDA FMD Vaccination Policy states the following:

The goal (of this Policy) is to advance preparedness by facilitating discussion, if not consensus, among our many partners to identify what level of preparedness is adequate and cost effective when considering:

- *Procuring and maintaining a sufficient amount of vaccine for a large-scale emergency vaccination effort is extremely costly.*

- *Vaccine quantity currently available to USDA is sufficient to respond to a small, focal outbreak in an area that is not livestock-dense.*
- *FMD virus strains are sufficiently different so vaccinating against one strain may not protect against different strains, even if they are related.*
- *FMD vaccine cannot be currently produced in the United States (21 U.S.C. 113A). The current vaccine antigen concentrate (VAC) held by the North American FMD Vaccine Bank must be shipped abroad to be finished into vaccine.*
- *VAC currently held by the North American FMD Vaccine Bank is intended to be shared by the United States, Canada, and Mexico. For VAC currently held by the North American FMD Vaccine Bank, the vaccine manufacturers can produce 2.5 million doses in 21 days upon receiving the VAC. For additional vaccine (created from a master seed and not currently stored as VAC), vaccine production can take as long as 14 weeks.*

In working with our stakeholders, USDA-APHIS believes that an efficient, overall approach to protect the Nation's livestock industry in an FMD outbreak can be developed. Although the vaccination aspect of preparedness presents unique challenges, these can be overcome with adequate advance planning and consideration of the capabilities and opportunities that public-private partnerships and cost-sharing can afford.

RESOLUTION:

The United States Animal Health Association urges the United States Secretary of Agriculture, in concert with the appropriate agencies, to include a request for funding in the Fiscal Year 2018 budget to develop an optimal Foot-and-Mouth Disease (FMD) Vaccine Bank and to create an FMD Preparedness and Response Plan that supports continuity of business within the United States animal agriculture industry should a large scale, multi-state, multi-strain FMD outbreak occur. The development of this budget should be informed by the criteria set forth in the United States Department of Agriculture (USDA) Sources Sought Notice (Solicitation Number: AG-6395-S-16-0086) issued by the USDA on March 14, 2016.

The request submitted should be adequate to fund expansion of existing FMD virus antigen stockpiles to allow for production of sufficient quantities of FMD vaccine by capable vaccine manufacturers to produce 25 million doses in a timely fashion of each of the top 10-13 FMD virus strains recommended by the FMD World Reference Laboratory (WRLFMD) for FMD vaccine banks in FMD-Free countries.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 5

APPROVED

SOURCE:

COMMITTEE ON ANIMAL EMERGENCY MANAGEMENT

SUBJECT MATTER:

Termination of the American Veterinary Medical Association's Veterinary Medical Assistance Team Program and Participation in Disaster and Emergency Response

BACKGROUND INFORMATION:

The American Veterinary Medical Association (AVMA) website provides this historical data: *"VMAT was founded in 1992 in the aftermath of Hurricane Andrew which caused significant damage in Florida and inflicted heavy losses on animals and the veterinary infrastructure. In 1993, the AVMA signed a Memorandum of Understanding (MOU) with the United States Department of Health and Human Services (HHS), making VMAT part of the Federal Response Plan (now the National Response Framework) as part of the National Disaster Medical System (NDMS). In 1994, the AVMA entered into an MOU with the United States Department of Agriculture, making VMAT available to respond in the event of an animal health emergency.*

Over the years, VMAT members provided on the ground veterinary support during a number of disasters and emergencies including the Hurricanes Katrina, Rita and Wilma in 2005 and the World Trade Center Attacks in 2001 as well as many other events.

In 2008 the federal law changed, and the public-private partnership was dissolved. This led to the creation of two distinct veterinary response programs: The National Veterinary Response Teams (NVRT), part of NDMS at HHS, and the AVMA's VMAT program. These organizations collaborate, communicate and cooperate with each other on issues related to animal emergency preparedness and response. MOUs between the AVMA and HHS signed in 2008 and 2012 highlight the relationship.

With the change in the federal law, VMAT's program evolved. The current VMAT program focuses on state-level response. AVMA VMAT teams are available to deploy at the request of the state to assist in animal emergency response and deploy within the state's incident command structure. VMAT has three missions: 1) Providing on-the-ground assessment of veterinary infrastructure following a disaster. Reports provided by VMAT volunteers in the field can be utilized by State emergency response officials to direct resources to impacted areas. 2) Augmenting state veterinary response resources to provide veterinary care to animals affected by a disaster. 3) Providing training on a wide range of veterinary disaster response topics to veterinary response

organizations, veterinary medical associations, veterinary students and other related organizations through the VMAT U program.”

In June of 2016 the AVMA Committee on Disasters and Emergency Issues (CDEI) were tasked with making recommendations to the AVMA Executive Board regarding re-organization of the VMAT teams in order for the board to have background material. Termination of the program was not recommended although changing from a response unit to a preparedness unit was recommended due to the large number of states and local jurisdictions that have their own animal response teams.

On September 22, 2016, VMAT team members and CDEI committee members were notified by the Chief Executive Officer of the AVMA, Dr. Janet Donlin, that the Executive Board was discontinuing the VMAT program over a 12-18 month period of time.

RESOLUTION:

The United States Animal Health Association (USAHA) recommends that the American Veterinary Medical Association (AVMA) Board of Directors reestablish the Veterinary Medical Assistance Team program as a veterinary educational resource for veterinarians assisting animals in disasters by providing training and exercises to help build state and local capacity, by providing incident support as subject matter experts and, finally, promoting business continuity planning and disaster recovery for veterinarians.

USAHA recommends that the AVMA Committee on Disasters and Emergency Issues continue to be an active partner in animal disaster preparedness.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 6, 13, 29, 34, and 42 Combined

APPROVED

SOURCE:

**COMMITTEE ON INFECTIOUS DISEASES OF CATTLE,
BISON AND CAMELIDS
COMMITTEE ON INFECTIOUS DISEASES OF HORSES
COMMITTEE ON CAPTIVE WILDLIFE AND ALTERNATIVE
LIVESTOCK
COMMITTEE ON TRANSMISSIBLE DISEASES OF
POULTRY AND OTHER AVIAN SPECIES
COMMITTEE ON SHEEP AND GOATS**

SUBJECT MATTER:

Laboratory Approval for Regulatory Diseases

BACKGROUND INFORMATION:

Laboratories performing animal surveillance testing are integral to establishing the health status of the national herds and flocks as well as individual animals destined for export. Currently, the United States Department of Agriculture has no authority to restrict laboratories from conducting foreign animal disease diagnostic testing on livestock and poultry samples. For example, one private laboratory in the United States is advertising Polymerase Chain Reaction (PCR) testing for all of the following Foreign Animal diseases African swine fever, African horse sickness, avian influenza, classical swine fever, foot and mouth disease, nipah virus, newcastle disease virus, pestes des petits ruminants virus, rinderpest, rift valley fever, contagious equine metritis, glanders, piroplasmiasis, and surra.

Additionally, there is limited state authority or oversight over laboratories conducting diagnostic testing for diseases of regulatory importance on samples obtained from livestock and poultry. The same laboratory conducting the above listed foreign animal disease tests offers PCR tests for equine infectious anemia, brucellosis, infectious bursal disease, influenza, Johnes disease, pseudorabies, Q fever, rabies, West Nile Virus and vesicular stomatitis. Other private laboratories are promoting new diagnostic testing modalities for diseases of regulatory importance such as chronic wasting disease.

There is no requirement that the tests offered by unregulated laboratories are approved and validated to accurately assess infection status, nor are there requirements that tests offered by unregulated laboratories conform to national regulatory testing requirements. Diagnostic tests, especially PCR, are difficult to perform and a small deviation from standards could potentially result in a false positive or false negative test result. The practitioner or producers are likely not aware of these difficulties in performing the test or the potential regulatory ramifications of a false test result reported to state animal health

officials. Ultimately, the inability to prescribe laboratory testing standards necessary for ensuring the health of livestock and poultry, places animal agriculture in the United States at significant risk.

RESOLUTION:

The United States Animal Health Association (USAHA) urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service, Veterinary Services to restrict foreign animal disease diagnostic testing to laboratories approved by the USDA and to take regulatory enforcement action against non-approved laboratories conducting testing for foreign animal diseases. If USDA doesn't currently have authority for these actions, USAHA urges USDA to take measures to establish those authorities.

Additionally, the USAHA recommends state animal health officials assess state authority or oversight over laboratories conducting diagnostic testing for diseases of regulatory importance on samples obtained from livestock and poultry.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 7, 9, 11 and 24 Combined

APPROVED

SOURCE:

**USAHA/AAVLD COMMITTEE ON ANIMAL HEALTH
SURVEILLANCE AND INFORMATION SYSTEMS
USAHA/AAVLD COMMITTEE ON NATIONAL ANIMAL
HEALTH LABORATORY NETWORK
COMMITTEE ON TRANSMISSIBLE DISEASES OF SWINE
COMMITTEE ON BIOLOGICS & BIOTECHNOLOGY**

SUBJECT MATTER:

**Sustained Fiscal Year 2017 Funding for the United
States Department of Agriculture, Animal and Plant
Health Inspection Service / Influenza A Virus – Swine
Surveillance Activities**

BACKGROUND INFORMATION:

Economic losses due to influenza A virus in swine (IAV-S) infections are substantial and a global problem, ranking among the top three major health challenges in the swine industry. In addition, IAV-S continues to be a concern to public health and the poultry industry.

The United States Department of Agriculture (USDA) began a surveillance system in 2009 to better characterize the genetic diversity of IAVs of swine. Data from this surveillance system have revealed tremendous genetic diversity across IAV-S isolates. This diversity creates great challenges for effective vaccination control programs. The need for next generation swine influenza vaccines that elicit broader cross-protection has never been greater.

The IAV-S Surveillance Program has collected and characterized virus isolates from swine since it was initiated in 2009. The program supports both animal and public health objectives. Program goals include monitoring the evolution of the virus, providing isolates for research and the development of diagnostic reagents, and updating diagnostic tests and vaccine Master Seed stocks. Importantly, the information gained from the surveillance system has benefitted our human health counterparts at the United States Department of Health and Human Services (HHS), Centers for Disease Control and Prevention (CDC) by providing sequence data from isolates identified in inter-species spillover events.

Following the human vaccine model, IAV-S vaccine backbones could be approved by the USDA, Animal and Plant Health Inspection Service (APHIS), Center for Veterinary

Biologics (CVB) to permit timely updates with new and relevant hemagglutinin (HA) and neuraminidase (NA) for vaccine seed strains. These backbones should exhibit high yield growth properties as well as attenuating mutations in the case of live attenuated influenza vaccine. Commercial vaccine manufacturers may select viruses based on HA and NA sequences from the surveillance system or from their own internal surveillance data for their customers. Viruses from the USDA IAV-S surveillance repository are readily available for this purpose.

CDC funds were provided for an initial pilot influenza surveillance project in 2008. Additional surveillance activities were funded by allocations from the HHS to USDA-APHIS as one-time, no year funds under the authority of the Supplemental Appropriations Act of 2009 for pandemic influenza preparedness and response. Those funds will run out in Fiscal Year 17. This surveillance system is the best system in the world and has contributed greatly to understanding the influenza status in swine in the US and provides an incredibly valuable public health resource for the CDC.

RESOLUTION:

The United States Animal Health Association requests the 115th United States Congress to appropriate and the Secretary of Agriculture to allocate a minimum of \$10 million of mandatory funding in future United States Department of Agriculture, Animal and Plant Health Inspection Service budgets for influenza A virus in swine surveillance as part of a comprehensive and integrated swine disease surveillance program.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 8

APPROVED

SOURCE:

AAVLD/USAHA COMMITTEE ON AQUACULTURE

SUBJECT MATTER:

**Quality Assurance Training for Aquatic Animal
Laboratories**

BACKGROUND INFORMATION:

Pathogen testing is a legal requirement for transport, export and management of aquatic animals in many jurisdictions within the United States and beyond. It is critical that the results of this testing be accurate, credible and beyond reproach. Currently, many laboratories performing this testing have little or no quality assurance/quality control programs in place and no training available. Many smaller state, federal or tribal laboratories feel they do not have the resources to accomplish accreditation through World Organization for Animal Health (OIE) or International Organization for Standardization (ISO) 17025.

A grassroots effort has been initiated through the Fish Health Section (FHS) of the American Fisheries Society (AFS) to create and implement a voluntary, multi-tiered approach for quality assurance, based largely on Chapter 3 of the FHS/United States Fish and Wildlife Services Blue Book, as well as key ingredients of other accreditation programs. A standing committee consisting of several state, federal and private partners has created the first Tier (pre-qualification) which was announced in January 2016. The second Tier (recognition) is under development. The committee is exploring ways to partner with other agencies with ongoing programs to help further this effort.

RESOLUTION:

The United States Animal Health Association encourages the United States Department of Agriculture, Animal and Plant Health Inspection Service and the National Animal Health Laboratory Network to provide support and initiate quality management training for the American Fisheries Society (AFS), Fish Health Section (FHS) Quality Assurance Initiative which can be provided at regional meetings of the aquatic diagnostic testing community (e.g., AFS-FHS annual meeting, Eastern Fish Health Workshop, and Western Fish Disease Workshop).



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 10

APPROVED

SOURCE: USAHA/AAVLD COMMITTEE ON NATIONAL ANIMAL HEALTH LABORATORY NETWORK

SUBJECT MATTER: Laboratory Requirements for Program Disease Testing

BACKGROUND INFORMATION:

Quality assurance for animal disease laboratory testing and electronic messaging of diagnostic results are of critical importance for maintaining and enhancing the state of preparedness for actively managing diseases of high consequence to United States (US) animal health. Quality assured test results and seamless (electronic) connectivity of information between diagnostic laboratories and the appropriate state and federal veterinary medical agencies are needs of 21st century US animal agriculture.

RESOLUTION:

The American Association of Veterinary Laboratory Diagnosticians (AAVLD) and the United States Animal Health Association (USAHA) recommend that a working group inclusive of representation from AAVLD, USAHA, the United States Department of Agriculture, and the United States Food and Drug Administration (FDA) be formed to provide a formal review and subsequent recommendation for implementation of minimum quality and reporting requirements of laboratories responsible for conducting diagnostic testing associated with determination of animal disease status under the various USDA and FDA programs. The resulting review and recommendations for consideration should be provided to each of the contributing organizations prior to the 2017 Annual Meeting of USAHA and AAVLD.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 12 and 14 Combined

APPROVED

**SOURCE: COMMITTEE ON TRANSMISSIBLE DISEASES OF SWINE
COMMITTEE ON INFECTIOUS DISEASES OF HORSES**

**SUBJECT MATTER: State Animal Health Official and Submitting Veterinary
Diagnostic Lab Access to Veterinary Diagnostic Laboratory
Records Reported from the National Animal Health Laboratory
Network Labs and the National Veterinary Services Laboratory
to the United States Department of Agriculture's Laboratory
Messaging Service**

BACKGROUND INFORMATION:

The United States Department of Agriculture's (USDA) Laboratory Messaging Service (LMS) is a database application that serves as the centralized point of receipt for electronic veterinary diagnostic records being reported from veterinary diagnostic labs (National Animal Health Laboratory Network (NAHLN) labs) to the USDA. LMS also receives test results being reported from cases forwarded from NAHLN labs to the USDA, National Veterinary Services Laboratory (NVSL) for further diagnostic testing. Significant advances have been made in the NAHLN's ability to electronically transfer (message) veterinary diagnostic records from NAHLN labs and NVSL to LMS. These stepwise improvements in connectivity between veterinary diagnostic laboratories (VDLs) and USDA represent great progress towards establishing seamless and scalable systems of reportable disease veterinary diagnostic information transfer between US VDLs and veterinary medical officials. However, USDA does not currently have an effective application for providing state animal health officials electronic access to the veterinary diagnostic laboratory records received into LMS that have originated from animals or farm sites in their respective states. Similarly, NAHLN labs do not have electronic access to diagnostic results from case submissions in which they forward onto NVSL for further testing. Permissioned access solutions are needed to bridge this gap in connectivity that exists between the USDA's LMS, state animal health officials, and veterinary diagnostic laboratories.

RESOLUTION:

The American Association of Veterinary Laboratory Diagnosticians (AAVLD) and the United States Animal Health Association (USAHA) encourage the United States Department of Agriculture (USDA) to develop an application that provides state animal health officials electronic access to veterinary diagnostic laboratory records originating from animals or farm sites in their respective states that have been reported from National Animal Health Laboratory Network Labs or USDA, National Veterinary Services Laboratory (NVSL) to USDA's Laboratory Messaging Service. Similarly, AAVLD and USAHA encourage USDA to provide veterinary diagnostic laboratories electronic access to diagnostic results from case submissions in which that same veterinary diagnostic laboratory has forwarded onto NVSL for further testing.



RESOLUTION NUMBER: 15 and 45 Combined

APPROVED

**SOURCE: COMMITTEE ON INFECTIOUS DISEASES OF HORSES
COMMITTEE ON PARASITIC AND VECTOR BORNE
DISEASES**

**SUBJECT MATTER: Equine Infectious Anemia and Equine Piroplasmosis
Testing of Racing Quarter Horses**

BACKGROUND INFORMATION:

Racing Quarter Horses have been identified as a high-risk population of horses which pose a significant risk to the health of the national equine population. Since 2009, there have been 268 racing Quarter Horses confirmed positive for equine piroplasmosis (EP), with 56 of the 268 confirmed since October of 2015. The 56 positive horses were located all across the country including in the states of Arkansas (2), Arizona (3), California (1), Illinois (1), New Mexico (1), North Carolina (1), Tennessee (19), Texas (10) and Wyoming (14). Additionally, since 2012, at least 59 racing Quarter Horses have been confirmed positive for equine infectious anemia in states of California (39), Texas (5), Washington (10), Oregon (4), and Oklahoma (1). Epidemiologic investigations into these cases have indicated iatrogenic transmission of disease through high risk practices of trainers and owners. The failure to promptly identify positive animals poses a significant risk to the United States (US) equine population as the retired racing Quarter Horses travel across the US to be used as pleasure horses, roping or rodeo horses, barrel horses, show horses or ranch horses. Of concern regarding equine piroplasmosis, the US free status is at risk if identification and control measures are not implemented. Although it is acknowledged that imposing testing requirements on racing Quarter Horses prior to entry into a racing venue will impose an increased owner expense, the threat of the loss of US free status for EP, and the threat of allowing permanent establishment of a new disease into the US horse industry poses an even greater economic risk to the US equine industries.

RESOLUTION:

The United States Animal Health Association (USAHA) urges state animal health officials and Quarter Horse racing jurisdictions to impose equine infectious anemia (EIA) and equine piroplasmosis (EP) testing requirements for Quarter Horses entering a racing venue. Additionally, USAHA urges the American Quarter Horse Association to encourage the EIA and EP testing of racing Quarter Horses and assist in the education of the racing Quarter Horse owners and trainers as to the risks of the diseases. Lastly, the USAHA urges the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services to continue to compile national epidemiologic EIA and EP data for the high-risk group of horses and provide outreach information to states and industry regarding this issue.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 16, 23, and 40 Combined

APPROVED

SOURCE:

**COMMITTEE ON SCRAPIE
COMMITTEE ON LIVESTOCK IDENTIFICATION
COMMITTEE ON SHEEP AND GOATS**

SUBJECT MATTER:

**Continued United States Department of Agriculture
Provision of Plastic Scrapie Program Ear Tags for Sheep
and Goats Producers**

BACKGROUND INFORMATION:

While the National Scrapie Eradication Program (NSEP) has been successful in decreasing the prevalence of scrapie in the United States, eradication has not yet been achieved in sheep or goats. Continued improvement in traceability and surveillance is needed, not just to achieve the eradication of scrapie, but also to advance animal disease traceability (ADT) efforts.

Much of the success of the NSEP is attributable to the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service's (APHIS) work with producers to find identification (ID) devices which have good retention and lend themselves to improving animal care and management. Currently, the USDA provides small metal tags or more visible plastic ear tags to producers, sales yards, fairs, veterinarians and veterinary clinics. The plastic tags have a larger profile and lend themselves to management systems where tag numbers are read and recorded. The metal tags are too small to be used as visible ID for management purposes, and they are more likely to lead to infections in goats than the plastic tags.

The publication of the NSEP final rule is expected in 2017 and will include new requirements for official identification and traceability for certain classes of goats and sheep previously excluded from mandatory official ID. In addition to the increasing numbers of new sheep and goat producers entering the program on a continuing basis, longtime producers of low risk goats and sheep, who were previously exempted, will have mandatory ID requirements for the first time. A change in tag-provision policy at this critical time jeopardizes the ability of veterinarians and scrapie program officials to facilitate compliance by these herd owners. Elimination of USDA-provided tags that provide best visible ID will compromise accurate recording of ID and compromise compliance with record keeping requirements for both traceability and the scrapie program

Alternative sources of funds and cost saving options to support the USDA-provided plastic ear tags should be explored. Benefits of the USDA-provided plastic tags outweigh the savings that could be achieved by cutting the funding for this item. The success in ADT attributable to the NSEP and the wide adoption of sheep and goat plastic ear tags demonstrate the value of providing ID options that benefit both producers and traceability.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service to continue to provide plastic ear tags for the National Scrapie Eradication Program (NSEP) in the most economical and case appropriate manner. These USDA-provided tags are critical to successful identification and traceability of sheep and goats for NSEP and animal disease traceability.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 17 and 41 Combined

APPROVED

SOURCE:

**COMMITTEE ON SCRAPIE
COMMITTEE ON SHEEP AND GOATS**

SUBJECT MATTER:

Goat Scrapie Genetic Resistance

BACKGROUND INFORMATION:

Genotype selection for scrapie resistance in sheep has proven to be a great asset in efforts to eradicate scrapie in sheep. The availability of genetic tools for goats should have similar benefits. Based on information presented by the United States Department of Agriculture, Agricultural Research Service researchers, sufficient data exists to support further efforts toward testing for goat scrapie genotype resistance and development of field applications in the National Scrapie Eradication Program.

RESOLUTION:

The United States Animal Health Association (USAHA) urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service to pursue efforts to develop pilot projects to explore the use of goat scrapie genotype testing in the National Scrapie Eradication Program. USAHA also requests that USDA-Agricultural Research Service conduct surveys to assess the frequency of resistant genotypes in United States (US) goats and identify methods to expand the availability of resistant genotypes to US goat producers.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 18

APPROVED

SOURCE:

COMMITTEE ON SCRAPIE

SUBJECT MATTER:

Identifying Non-Traditional Sheep and Goat Marketing and Slaughter Channels

BACKGROUND INFORMATION:

While the National Scrapie Eradication Program (NSEP) has been successful in decreasing the prevalence of scrapie in the United States, eradication has not yet been achieved. With all disease eradication programs, as prevalence of the disease declines the ability to identify the remaining cases becomes an ever greater challenge.

There is evidence that increasing numbers of sheep and goats are marketed and slaughtered outside of the traditional marketing system and may not be available for scrapie surveillance, the impact of which may prolong the time until eradication is achieved. It is also likely that the demand for nontraditionally marketed animals will continue to rise resulting in negative ramifications for the program.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services to actively pursue identifying nontraditional sheep and goat marketing and slaughter channels and to create a program to obtain samples from these channels.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 19

APPROVED

SOURCE:

COMMITTEE ON BRUCELLOSIS

SUBJECT MATTER:

Review of State Brucellosis Management Plans

BACKGROUND INFORMATION:

All states are essentially free of bovine brucellosis, and the disease has largely been eliminated from the United States (US) cattle population, despite occasional 'spillover' infection from infected wildlife reservoirs. The United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service (APHIS) published an Interim Rule in 2010, which effectively removed state status for bovine brucellosis. Eleven infected cattle or domestic bison herds have been detected within the Designated Surveillance Area (DSA) as a result of the testing required within the DSA since 2010, effectively identifying and mitigating disease risk to the US cattle population.

Under the 2010 Interim Rule, the Greater Yellowstone area (GYA) states are responsible for defining the boundaries of the DSA, conducting surveillance "sufficient to prevent the spread of brucellosis...", and implementing a Brucellosis Management Plan (BMP), approved by USDA-APHIS, Veterinary Services (VS) in a Memorandum of Understanding (MOU). USDA-APHIS-VS last reviewed GYA state BMPs in 2012.

State required testing of DSA cattle and domestic bison herds appears to be effective in identifying infected herds at low prevalence. Affected herds are being identified prior to leaving the DSA, no herds have been found infected outside of the DSA, and no cases of herd-to-herd transmission have been documented since the 2010 rule and implementation of DSA required testing.

However, surveillance in wildlife outside of the Wyoming DSA has identified seropositive elk annually for the last four years, and the boundaries of the DSA have not been expanded accordingly. The finding of seropositive elk in areas outside of a DSA may indicate current or past infection, the implication of which is that cattle and domestic bison herds in the area may also be at risk of infection. Lack of timely action in expanding DSA boundaries in response to finding exposed wildlife may result in exposed or infected cattle or bison leaving the area undetected.

RESOLUTION:

The United States Animal Health Association (USAHA) urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service (APHIS)

to conduct reviews of Greater Yellowstone Area (GYA) state Brucellosis Management Plans and their implementation, at least once every three years. In addition, USAHA also encourages GYA states and USDA-APHIS to continue to conduct wildlife surveillance outside of Designated Surveillance Areas (DSA), and for the states to adjust DSA boundaries accordingly to include geographic areas where there is a potential risk of transmission of brucellosis from wildlife to cattle or domestic bison.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 20

APPROVED

SOURCE:

COMMITTEE ON BRUCELLOSIS

SUBJECT MATTER:

**Brucellosis Milk Enzyme Linked Immunosorbent Assay
Validation as an Additional Test for Brucellosis in Bulk
Milk**

BACKGROUND INFORMATION:

At present, the Brucellosis Ring Test (BRT) is the only approved test for detection of antibodies to *Brucella spp.* in bulk milk tank samples, but this test consistently demonstrates false positives. If the brucellosis milk enzyme linked immunosorbent assay (ELISA) is demonstrated to have improved sensitivity and specificity, the United States Animal Health Association supports work by the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services to validate the ELISA, in addition to the BRT, as an approved brucellosis bulk milk surveillance test.

The IDEXX milk ELISA test was available for bulk milk tank sampling from the early 2000s until 2006 or 2007. During that time, the milk ELISA was utilized for bulk milk tank samples due to the superior sensitivity when compared to the BRT. When IDEXX discontinued the production of the milk ELISA test, the BRT was the only approved test for bulk milk tank sampling in the United States.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service to direct the National Veterinary Services Laboratory to pursue validation of the brucellosis milk enzyme linked immunosorbent assay.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services to amend the live cervid import protocol upon request from the Canadian Food Inspection Agency to exclude Manitoba cervids that originate outside the Riding Mountain National Park Tuberculosis Eradication Area from the isolation requirements prior to testing.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 25

APPROVED

SOURCE:

COMMITTEE ON BIOLOGICS & BIOTECHNOLOGY

SUBJECT MATTER:

International Promotion of the United States Regulatory System for the Regulation of Veterinary Biological Products

BACKGROUND INFORMATION:

The United States Department of Agriculture (USDA) has regulated the veterinary biologics industry for over 100 years, and under the Virus-Serum-Toxin Act of 1913 (amended in 1985), has developed regulatory methods to ensure that the veterinary biologics manufactured in the United States (US) are pure, safe, potent, and effective. This regulatory system is described in the Code of Federal Regulations (CFR) beginning at 9 CFR part 101 (Subchapter E). In 2015, the US domestic veterinary biologics industry manufactured over 100 billion doses of high quality products for both domestic and global markets. More recently, certain countries (often with local industry support interests) have intimated in the international arena that the US regulatory system is not equivalent, and by implication inferior, to their regulatory systems. Consequently, some countries are no longer accepting inspection certification from the USDA. Some insist on conducting their own inspections, while others will accept certificates of inspection from a regulatory body that is a member of the Pharmaceutical Inspection Cooperation Scheme (PIC/S). PIC/S is a non-binding cooperative arrangement between regulatory authorities to promote quality inspections and to facilitate cooperation and networking among these authorities to promote mutual confidence. Approximately fifty regulatory authorities are currently members of the PIC/S.

The USDA, Animal and Plant Health Inspection Service (APHIS), Center for Veterinary Biologics has worked to promote an understanding of their regulatory system through participation in the Veterinary International Conference on Harmonization (VICH), outreach to Latin America through a World Organization of Animal Health (OIE) program called CAMEVET, and participation in the Institute of International Cooperation in Animal Biologics, which is an OIE collaborating center located at Iowa State University that conducts programs that are often attended by foreign governments. These outreach efforts are appreciated, however more is needed to help promote and protect export markets for US veterinary biologics. Countries with recent specific issues include Russia, Thailand, and Turkey. The USDA-APHIS-CVB should evaluate leveraging other ongoing USDA-APHIS trade outreach programs to further promote their regulatory system and evaluate joining the PIC/S.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service programs, including International Services, Veterinary Services' National Import Export Services, and Center for Veterinary Biologics to develop a plan to increase USDA efforts to promote the United States regulatory system for veterinary biologics as a high quality regulatory system designed to ensure the production of pure, safe, potent, and efficacious veterinary biological products. The plan should specifically evaluate outreach to problematic areas and joining the Pharmaceutical Inspection Cooperation Scheme.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 27

APPROVED

SOURCE:

COMMITTEE ON PUBLIC HEALTH AND RABIES

SUBJECT MATTER:

Increased Fiscal Year 2018 Funding for the United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services Oral Rabies Vaccination Program

BACKGROUND INFORMATION:

The United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service (APHIS), Wildlife Services (WS), National Rabies Management Program (NRMP) has demonstrated through the strategic implementation of cooperative oral rabies vaccination (ORV) programs targeting wildlife to be cost-effective, while continuing to reduce rabies exposure and transmission among wildlife, livestock, pets and people. The United States Animal Health Association agrees with the World Organization for Animal Health (OIE); the most effective strategy to implement large scale rabies control efforts is at the source in animal (i.e., vector) populations. ORV programs are designed to immunize target wildlife species by increasing the percentage of rabies-immune animals within vaccination zones. Creating a population of immune animals results in the reduction of rabies cases, prevention of viral spread, and eventual rabies elimination.

In early 2016, WS with federal, state, academic, and international experts developed a comprehensive strategy to implement Phase 2, elimination of raccoon rabies variant in the Eastern United States. WS also developed and initiated an Enhanced Rabies Surveillance Program with state cooperation throughout the Northeast, Atlantic, and adjacent Mid-West and Southern States to enhance early detection of rabies cases or translocation of animals with rabies. This will allow for rapid contingency plans to eliminate rabies from re-infected areas and minimize the threat of rabies spread to newly infected areas.

Successful programs in Texas continue towards rabies elimination in gray foxes, as well as ongoing studies on rabies control methodology in skunks and maintaining a protective immune barrier along the Mexican border to keep the United States free of coyote (canine) rabies and **prevent having gray fox rabies elimination efforts undermined by entry of rabid foxes into Texas from Mexico**. The requested funding will allow USDA to:

- Fully implement the enhanced rabies surveillance program.
- Implement contingency action in response to rabid animals in sensitive areas.

- Continue Phase 1 as outlined in the US National Plan for Wildlife Rabies Management that maintains existing operational programs (immune zones) to control rabies in wildlife populations.
- Continue the investigation of novel and US-licensed vaccines and baits.
- Continue studies related to rabies control in skunks.
- Initiate Phase 2 of the national plan to eliminate raccoon rabies variant in the U.S.

RESOLUTION:

The United States Animal Health Association requests the 115th Congress to appropriate a minimum of \$30 million for program management and contingency actions at the state level in the Fiscal Year 2018 budget line item for the United States Department of Agriculture, Animal and Plant Health Inspection Service, Wildlife Services National Rabies Management Program.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 28

APPROVED

SOURCE:

COMMITTEE ON PUBLIC HEALTH AND RABIES

SUBJECT MATTER:

Wildlife Translocation: A Threat to Rabies Management and Elimination Programs

BACKGROUND INFORMATION:

Intentional and accidental translocation of meso-carnivores can result in significant management challenges and seriously threatens wildlife rabies management initiatives being implemented by the Wildlife Services (WS) National Rabies Management Program (NRMP) and its international, national, and state partners. Translocation often occurs as a response to human-wildlife conflicts. The intentional relocation of raccoons, skunks, foxes, and coyotes is typically carried out by the public, nuisance wildlife control operators, wildlife rehabilitators, and others to move nuisance or rehabilitated animals away from the site of capture. Accidental translocation (hitch-hikers) by vehicle, boat, or airplane may result in local, interstate, or international movement that can facilitate the spread of rabies. Raccoon rabies was documented again in Ontario Canada last fall after more than 10 years of being raccoon rabies free. The Ontario Ministry of Natural Resources immediately implemented a contingency action plan to control, minimize, and work to eliminate the outbreak. The Canadian Food Inspection Agency in Ottawa, Canada determined by DNA sequence of the raccoon rabies isolate that the likely origin was from southeastern New York State or New York City and it probably reached Ontario by translocation. Recently, several confirmed raccoon translocation events occurred in states where oral rabies vaccination (ORV) programs were being conducted to stop the spread of raccoon rabies, underscoring potential impacts on broad scale rabies management programs in 15 states.

State fish and wildlife agencies have legal jurisdiction and management authority over the common rabies vector species (RVS) in the United States. The South Eastern Association of Fish and Wildlife Agencies Fur Working Group (FWG) developed recommendations for their respective state agencies that are attempting to prevent the spread of rabies by RVS. The FWG developed Best Management Practices (BMP) for various user groups, wildlife damage control agencies, wildlife rehabilitators, trappers, and the general public that may encounter RVS. The BMPs are sound and work on the basis that no RVS should be relocated or translocated, but instead should be released at the capture site or humanely euthanized. Many states allow user groups to release RVS away from the original site of capture and state laws and regulations vary greatly on the legality of transporting across state lines.

Translocation of RVS can result in significant costs to cooperative ORV programs that have to implement contingency actions to reestablish raccoon rabies free areas once raccoon rabies outbreaks occur. The development and implementation of a comprehensive public education strategy in concert with aggressive enforcement of state and local regulations prohibiting translocation of meso-carnivores is essential to reduce the economic burden of translocation of RVS.

RESOLUTION:

The United States Animal Health Association requests that the United States Department of Agriculture, Wildlife Services collaborate with local, state, and international partners to promote and, where legal and practical, implement the Best Management Practices for common rabies vector species developed by the South Eastern Association of Fish and Wildlife Agencies.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 30

APPROVED

SOURCE:

**COMMITTEE ON CAPTIVE WILDLIFE AND
ALTERNATIVE LIVESTOCK**

SUBJECT MATTER:

Live Animal Testing for Chronic Wasting Disease

BACKGROUND INFORMATION:

Detection of Chronic Wasting Disease (CWD) in live animals is an important component of CWD Prevention and Control Programs.

With the funding decrease for CWD indemnification, the need for a successful live animal test option, with a high rate of sensitivity and specificity, is critical in both a trace-forward / trace-back scenario, as well as in herd management plans.

There have been numerous studies evaluating the sensitivity and specificity of tonsillar biopsies in cervids. Similar to scrapie, PrP(CWD) in deer accumulates in the retropharyngeal lymph nodes and tonsillar follicles before central nervous system involvement or clinical symptoms (Sigurdson et al., 1999; Spraker et al., 2002b; O'Rourke et al., 2003). Antemortem testing of these tissues by immunohistochemistry provides a reliable preclinical diagnosis in deer (Wild et al., 2002; Wolfe et al., 2002).

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services to expedite evaluation and approval of tonsillar biopsies into the Chronic Wasting Disease (CWD) Program Standards, providing for rapid implementation and deployment as a viable, accurate, and reliable means of live animal testing for CWD in cervids.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

Resolution number: 31 and 39 Combined **APPROVED**

Source: COMMITTEE ON CAPTIVE WILDLIFE & ALTERNATIVE LIVESTOCK
COMMITTEE ON TUBERCULOSIS

Subject Matter: National Cervid Tuberculosis Herd Accreditation Program

BACKGROUND INFORMATION:

The primary objective of the cervid bovine tuberculosis (bTB) herd accreditation program is to eliminate *Mycobacterium bovis*, the causative agent of bTB, in farmed/captive cervids as part of a comprehensive approach to eradicate bTB in domestic cattle and bison in the United States. All farmed/captive cervids destined for interstate movement are required to be tested for bTB.

In 2005 Code of Federal Regulations (CFR) 9 Part 77 was updated to separate cervids from the cattle and bison program, and a new testing criteria for cervids was implemented. Herds that participate in the United States Department of Agriculture, Animal and Plant Health Inspection Service Cervid bTB Herd Accreditation Program must test their entire herd of cervids over 12 months of age, negative for bTB two times in 9 to 15 month intervals to establish an Accredited Free herd. The accreditation is valid for 33 to 39 months from the original anniversary date and a negative whole herd retest must be performed in that period of time to maintain the accredited status. Animals from the Accredited Free herds are allowed to be moved interstate at any time without further testing

Details on the bTB testing requirements for interstate movements of cervids from monitored herds, qualified herds, and accredited herds from modified accredited States and zones are provided in the federal regulations (9 CFR Parts 77 and 86) and in the 1999 UM&R on Bovine Tuberculosis Eradication.

Language from USDA Website referencing 1999 UM&R -

Bovine Tuberculosis (bTB) Testing Requirements for Interstate or International Movement

Last Modified: Apr 7, 2015

According to the 1999 TB UM&R:

1. No captive cervid with a response to any tuberculosis test is eligible for international movement.

2. No captive cervid with a response to any tuberculosis test is eligible for interstate movement unless said animal is subsequently classified “negative for tuberculosis” based upon an official tuberculosis test or is consigned directly to slaughter.
3. Captive cervids that originate from accredited herds may be moved interstate without further tuberculosis testing, provided that they are accompanied by a certificate stating that such captive cervids originated from an accredited herd.
4. Captive cervids not known to be affected with or exposed to tuberculosis that originate from qualified herds may be moved interstate if the animals are accompanied by a certificate stating that they originate from a qualified herd and have been classified negative to an official tuberculosis test that was conducted within 90 days prior to the date of movement. If the qualifying test was administered within 90 days of movement, the animal(s) to be moved do not require an additional test.
5. Captive cervids not known to be affected with or exposed to tuberculosis that originate from monitored herds may be moved interstate if they are accompanied by a certificate stating that such captive cervids originate from a monitored herd and have been classified negative to an official tuberculosis test that was conducted within 90 days prior to the date of movement.
6. Captive cervids not known to be affected with or exposed to tuberculosis that originate from all other herds may be moved interstate, provided that (1) they are accompanied by a certificate stating that such captive cervids have been classified negative in response to two official tuberculosis tests conducted no less than 90 days apart, (2) the second test was conducted within 90 days prior to the date of movement, and (3) the animals were isolated from all other members of the herd during the testing period.
7. Captive cervids less than 12 months of age that originate from and were born in qualified or monitored herds may be moved without further tuberculosis testing, provided that they are accompanied by a certificate stating that such captive cervids originated from such herds and have not been exposed to captive cervids from a lower status herd.
8. Institutions that have been accredited by the American Zoo and Aquarium Association (AZA) are exempt from these requirements when movement is between accredited member facilities. Captive cervids in zoological parks that have been accredited by AZA are exempt from the regulations in this subpart when the captive cervids are moved directly interstate between AZA member facilities. Any captive cervids moved interstate that are not moved directly from an AZA member facility to another AZA member facility must be moved in accordance with the regulations in this subpart.
9. Except for captive cervids moving interstate under permit directly to slaughter or necropsy, each captive cervid or shipment of captive cervids to be moved interstate must be accompanied by a certificate issued within 30 days of the movement by a State or Federal animal health official or an accredited veterinarian. The certificate must state the number of the official eartag or other identification approved by the Administrator for each captive cervid to be moved, the number of captive cervids covered by the certificate, the purpose of the

movement, the origin and destination of the captive cervids, the consignor, and the consignee.

Language from 1999 UM&R -

Part VI—Herd Status Plans for Captive Cervids

A. Accredited herd plan for captive cervids

1. Animals to be tested—Testing of herds for accreditation or reaccreditation shall include all captive cervids and all other hoof stock over 12 months of age and animals under 12 months of age that are not natural additions, except that animals under 12 months of age that are not natural additions originating from an accredited herd need not be tested.
2. Qualifying standards—To meet the requirements for accredited herd status, the herd must pass at least three consecutive official tests for tuberculosis conducted at 9- to 15-month intervals with no evidence of bovine tuberculosis.

In herds previously infected, the fourth, fifth, and sixth annual whole-herd negative test will requalify the herd for accreditation.

Herds meeting these standards may be issued a certificate by local State and Federal animal health officials.

3. Additions—Accredited herd additions must originate directly from one of the following and have no exposure to captive cervids from herds of lesser status than the additions' herd of origin:
 - a. An accredited herd.
 - b. A qualified or monitored herd, provided that the individual animals for addition had negative results on an official tuberculosis test conducted within 90 days prior to entry and were isolated from members of the accredited herd until these animals had a negative result on an official tuberculosis test conducted at least 90 days following entry.
 - c. A herd not meeting the requirements of (a) or (b) in this section. Individual animals for addition must be isolated from all other members of the herd of origin and must have negative results on two official tests for tuberculosis conducted at least 90 days apart. The second of these tests must be conducted within 90 days prior to movement to the premises of the accredited herd. The additions must be kept in isolation from members of the accredited herd until the additions have a negative result on an official tuberculosis test conducted at least 90 days following the date of entry.

Animals other than natural additions added to an accredited-free herd shall not receive the accredited herd status for sale or movement purposes until they have a negative result on a retest 90 days after entry and until they have been included in a recertification herd test.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services (VS) to issue a VS Guidance Document stating that “Animals other than natural additions added to an accredited-free herd shall not receive the accredited herd status for sale or movement purposes until they have a negative result on a retest 90 days after entry and until they have been included in a recertification herd test” is no longer applicable in the National Cervid Tuberculosis (TB) Herd Accreditation Program and no additional TB test is required for the accredited individual animal addition(s).



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 32

APPROVED

SOURCE:

COMMITTEE ON CAPTIVE WILDLIFE & ALTERNATIVE LIVESTOCK

SUBJECT MATTER:

Chronic Wasting Disease Testing Protocol for Wild Cervidae

BACKGROUND INFORMATION:

Over the last 15 years the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service (APHIS), Veterinary Services (VS) and state regulatory officials have worked to control and prevent the spread of Chronic Wasting Disease (CWD).

Producers raising CWD susceptible species can only move their animals interstate if they are in compliance with the CWD program set forth in Title 9 Code of Federal Regulations (CFR) Parts 55 & 81 that state animals must originate from herds with at least five years of CWD monitored status.

State wildlife agencies that plan and execute elk restoration projects from one state to another are moving CWD susceptible species interstate without following minimum interstate movement requirements for farmed cervidae. Instead, Title 9 CFR Part 81.3 states the source population be considered "low risk" by the receiving state and USDA APHIS.

To date, over two dozen herds of wild elk have been captured and transported to other states across the nation that follow no CWD protocol set forth in the CWD Program Standards.

The movement of CWD susceptible cervid species with unknown CWD status by state wildlife agencies can undermine the success of CWD control programs that have been in place in many states for more than 15 years. CWD has been found in 23 states. Eight of the 23 states have detected CWD in the free-ranging deer populations but not in the farmed cervid herds.

The USAHA Committee on Wildlife Diseases approved a resolution at the 2015 annual conference that requested USDA Veterinary Services to develop a guidance document for captive deer, elk, or moose captured from a wild population for interstate movement and release.

APHIS has finalized and released VS Guidance Document 8000.1 “Surveillance and Testing Requirements for Interstate Transport of Wild Caught Cervids” in October 2016 but the requirement of an ante-mortem test, such as the rectal biopsy, is only optional.

Exact language is as follows:

“Optionally, a whole-herd rectal biopsy or other mutually agreed-on method of antemortem CWD test with concurrent genotyping may be performed on the assembled herd. Laboratory results must be “not detected” on all animals. Animals with untestable or incorrect location samples (i.e., samples that are autolyzed or of the wrong tissue type) may be retested.”

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services (VS) to amend the language in VS Guidance Document 8000.1 “Surveillance and Testing Requirements for Interstate Transport of Wild Caught Cervids”, the Chronic Wasting Disease Program Standards, and Title 9 Code of Federal Regulations (CFR) Part 81.3, *(b) Animals captured for interstate movement and release*, to indicate that any wild cervid of a Chronic Wasting Disease (CWD) susceptible species captured and transported interstate for release shall require:

- 1) A rectal biopsy or other mutually agreed-on method of antemortem CWD test with concurrent genotyping performed on the assembled herd; and
- 2) Documentation of a sampling scheme sufficient to detect CWD at 1 percent prevalence with 95 percent confidence in wild cervids within the defined source population from which the animals are being moved and conducted within the most recent three-year period. Such sampling scheme shall include both passive (hunter harvest and found dead) and targeted surveillance for CWD.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 33

APPROVED

**SOURCE: COMMITTEE ON TRANSMISSIBLE DISEASES OF POULTRY
AND OTHER AVIAN SPECIES**

**SUBJECT MATTER: Approval of Real Time Reverse Transcriptase Polymerase
Chain Reaction Matrix Assay for Avian Influenza Surveillance
in National Poultry Improvement Plan Authorized Laboratories**

BACKGROUND INFORMATION:

National Poultry Improvement Plan (NPIP) authorized labs have successfully conducted avian influenza (AI) screening of flocks using agar gel immunodiffusion (AGID) and enzyme linked immunosorbent assay (ELISA) tests with approval of their Official State Agency (OSA), and state animal health officials for 18 years. The real time reverse transcriptase polymerase chain reaction (RRT-PCR) matrix assay for influenza A provides highly sensitive detection that is critical to ensure that birds are negative prior to translocation to other facilities. Authorized laboratories have successfully utilized molecular diagnostics for *Salmonella* and *Mycoplasma* and these assays have proven invaluable in NPIP program testing and compliance. An NPIP authorized primary breeder company laboratory not affiliated with the National Animal Health Laboratory Network (NAHLN) that uses a United States Department of Agriculture approved influenza A matrix assay RRT-PCR, achieves ISO 17025 quality certification, satisfactorily passes an National Veterinary Services Laboratory (NVSL) avian influenza matrix RRT-PCR proficiency test, and has an agreed memorandum of understanding (MOU) with their state animal health officials and official state agency (OSA) should be allowed to use the assay as a screening test within the NPIP's US Avian Influenza Clean program. Any non-negative detection at NPIP authorized laboratory would immediately be forwarded to the NVSL for confirmation and notification of state animal health officials and the OSA will occur as outlined in NPIP provisions and State animal health emergency protocols.

The following proposal was approved at the 2016 NPIP Biennial Conference:

§ 145.14 Testing

(d) For avian influenza.

(2) *Agent detection tests.* Agent detection tests may be used to detect influenza A matrix gene or protein but not to determine hemagglutinin or neuraminidase subtypes. Samples for agent detection testing should be collected from naturally occurring flock mortality or clinically ill birds.

(i) *The real time reverse transcriptase/polymerase chain reaction (RRT-PCR) assay.*

(A) The RRT-PCR tests must be conducted using reagents approved by the Department and the Official State Agency. The RRT-PCR must be conducted using the National Veterinary

Services Laboratories (NVSL) official protocol for RRT-PCR or a test kit licensed by the Department and approved by the OSA and the State Animal Health Official, and must be conducted by personnel who have passed an NVSL proficiency test.

- (a) For non-National Animal Health Laboratory Network (NAHLN) Authorized Laboratories:
- (i) RRT-PCR testing can only be used by primary breeder company Authorized Laboratories,
 - (ii) RRT-PCR testing can only be performed on their own breeding flocks and only used for routine surveillance,
 - (iii) the Authorized Laboratory has a quality system that is accredited as ISO/IEC 17025 or equivalent to perform the avian influenza RRT-PCR assay,
 - (iv) the Authorized Laboratory Memorandum of Understanding (MOU) included approval of use between the Authorized Laboratory, the Official State Agency (OSA), and the State Animal Health Official(s) of both the location of the Authorized Laboratory and the location where the breeder flocks reside,
 - (v) split samples for testing must occur between the Authorized Laboratory and a NAHLN laboratory at a frequency designated in the MOU.
- (B) Positive results from the RRT-PCR must be further tested by Federal Reference Laboratories using appropriate tests for confirmation. Final judgment may be based upon further sampling and appropriate tests for confirmation.

§ 146.13 Testing

(b) Avian influenza.

(2) *Agent detection tests.* Agent detection tests may be used to detect influenza A matrix gene or protein but not to determine hemagglutinin or neuraminidase subtypes. Samples for this testing should be collected from naturally occurring flock mortality or clinically ill birds.

(i) *The real time reverse transcriptase/polymerase chain reaction (RRT-PCR) assay.*

(A) The RRT-PCR tests must be conducted using reagents approved by the Department and the Official State Agency. The RRT-PCR must be conducted using the National Veterinary Services (NVSL) official protocol for RRT-PCR or a test kit licensed by the Department and approved by the OSA and the State Animal Health Official, and must be conducted by personnel who have passed an NVSL proficiency test.

- (a) For non-National Animal Health Laboratory Network (NAHLN) Authorized Laboratories:
- (i) RRT-PCR testing can only be used by primary breeder company Authorized Laboratories,
 - (ii) RRT-PCR testing can only be performed on their own breeding flocks and only used for routine surveillance,
 - (iii) the Authorized Laboratory has a quality system that is accredited as ISO/IEC 17025 or equivalent to perform the avian influenza RRT-PCR assay,
 - (iv) the Authorized Laboratory Memorandum of Understanding (MOU) included approval of use between the Authorized Laboratory, the Official State Agency (OSA), and the State Animal Health Official(s) of both the location of the Authorized Laboratory and the location where the breeder flocks reside,
 - (v) split samples for testing must occur between the Authorized Laboratory and a NAHLN laboratory at a frequency designated in the MOU.
- (B) Positive results from the RRT-PCR must be further tested by Federal Reference Laboratories using appropriate tests for confirmation. Final judgment may be based upon further sampling and appropriate tests for confirmation.

RESOLUTION:

The United States Animal Health Association (USAHA) urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service, Veterinary Services to approve the use of a USDA approved real time reverse transcriptase polymerase chain reaction matrix assay for influenza A in National Poultry Improvement Plan (NPIP) authorized primary breeder company laboratories as outlined in the NPIP proposed and passed change to the 9 Code of Federal Regulations 145.14 and 146.13 (Testing).



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 35

APPROVED

**SOURCE: COMMITTEE ON TRANSMISSIBLE DISEASES OF POULTRY
AND OTHER AVIAN SPECIES**

SUBJECT MATTER: Upland Gamebird Secure Poultry Supply Plan

BACKGROUND INFORMATION:

The upland gamebird industry is a \$1.9 billion industry that produces pheasants, bobwhite quail, chukar and Hungarian partridges for the United States gamebird hunting industry.

To minimize business interruption during a highly pathogenic avian influenza (HPAI) event, Secure Poultry Supply (SPS) plans for the table-egg layer, broiler and turkey industries are continuing to be developed using new risk assessments and past experience to act as tools to help emergency decision makers to provide rapid science-and risk-based decisions on the issuance or denial of movement permits within a Control Area.

The North American Gamebird Association, through its gamebird secure poultry supply plan working group, is attempting to develop a SPS plan for upland gamebirds that will be based upon a specific science- and risk-based plan that will include completed risk assessments.

RESOLUTION:

The United States Animal Health Association supports the current funding from United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services for the Upland Gamebird Secure Poultry Supply Plan risk assessments and encourages continued funding for these risk assessments beyond the current cooperative agreement.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 36

APPROVED

SOURCE: USAHA/AAVLD COMMITTEE ON DIAGNOSTIC LABORATORY AND VETERINARY WORKFORCE DEVELOPMENT

SUBJECT MATTER: Veterinary Public Practice Awareness and Promotion

BACKGROUND INFORMATION:

There have been several workforce studies over the last few years addressing the future of veterinary medicine and the critical role the profession plays in meeting societal needs, and the additional challenges the profession faces such as increased student debt. Most citizens of the nation are not aware of all the significant contributions veterinarians make to public health. To meet the increasing costs of veterinary education and the decreasing federal and state funding to support that education, veterinary colleges are increasing tuition and increasing class sizes in an attempt to meet those financial challenges.

A National Academy of Sciences (NAS) report from 2013 entitled “Workforce Needs in Veterinary Medicine” states that most of those students will likely practice companion animal medicine, and that “these actions will increase the supply of companion animal practitioners, the largest group of veterinary practitioners, at a time of uncertain demand for companion animal services”. The report further states that “the veterinary profession should expand its capacity to address complex global problems, such as those associated with food security, by encouraging interactions between US veterinary graduates and other disciplines and cultures, particularly in the developing world, where the profession has the opportunity to leverage its expertise in One Health and lead advances in food animal husbandry welfare, water safety and security, and the health of wildlife and ecosystems”. However, society must be convinced that investment in veterinary medicine is imperative. The study states that “the public, policymakers, and even medical professionals are frequently unaware of how veterinary medicine fundamentally supports both animal and human health and well-being” and that “broadening the public’s understanding will require commitment by veterinary leadership, the academe, and practitioners to develop and promote the profession as one that offers diverse career paths with many different niches for veterinarians, ranging from traditional companion animal practice to public and private sector positions in biomedicine, animal research, wildlife, the environment, global food production, food safety and security, and public health”.

An American Academy of Veterinary Medical Colleges (AAVMC) report of 2008 stated, “To safeguard the US economy, public health, and food supply, there must be recruitment and preparation of additional veterinarians into careers in public health, food systems, biomedical research, diagnostic laboratory investigation, pathology, epidemiology, ecosystem health, and food animal practice”. Conclusion 1 of the NAS report states in part “societal needs for veterinary expertise are substantial and growing, but the potential contributions of veterinary medicine are not realized because appropriate positions in relevant sectors are lacking.” Although there are many reasons why there has not been adequate public sector financial support of veterinary education

and opportunities, one clear reason is the lack of awareness of the public and decision-makers, and indeed many early career veterinary students, as to the value, skills, and broad interdisciplinary capabilities of veterinarians. To enhance the ability of the veterinary profession to better meet societal needs and to provide more opportunities for employment for veterinarians, it is critically important to increase public awareness of the skills, abilities, and broad-based training of veterinarians.

RESOLUTION:

The United States Animal Health Association urges the American Veterinary Medical Association to lead a public relations campaign similar to the “Partners for Healthy Pets” campaign, with a goal to raise public awareness of the breadth of skills of veterinarians in diagnostic and regulatory medicine and the contribution of veterinary medicine to public, animal and environmental health. Such a campaign could be called “Partners for a Healthy Planet”, “Partners for a Healthy Society,” or some such similar title.



RESOLUTION NUMBER: 38 **APPROVED**

SOURCE: **COMMITTEE ON TUBERCULOSIS**

SUBJECT MATTER: **Optimization and Standardization of Purified Protein Derivative Tuberculin Application for Interferon-gamma Release Assays**

BACKGROUND INFORMATION:

Infection with *Mycobacterium bovis* (*M. bovis*) continues to impact the United States cattle industry with a significant number of tuberculosis (TB) infected herds detected in different states in 2016. The caudal fold tuberculin (CFT) test is the primary screening test used in the bovine TB program. A major disadvantage of this test is that it requires cattle to be handled twice, once for the injection and a second time to interpret the test. Further, the person performing the test must also be adequately trained and sufficiently experienced to interpret the test results accurately. Experience is critical; determining a “response” may be subjective, especially if the response to the injection is weak. Test result accuracy may also depend on the purified protein derivative (PPD) tuberculin which is applied. Current regulation allows a range of potency as prescribed in the respective regulation of Title 9 Code of Federal Regulations (CFR) 113.409(c).

Currently used antibody tests demonstrate poor specificity resulting in too many false negative test results leading to undetected reactors remaining in the herd. In addition, antibody test can interfere with the caudal fold test.

BOVIGAM™ is one official auxiliary test used in cattle herds with the approval of the State Animal Health Official and United States Department of Agriculture, Animal and Plant Health Inspection Service, Veterinary Services, National Import Export Services Service Centers. This test may be used under direction of the Designated TB Epidemiologist and with concurrence of the Regional TB Epidemiologist. BOVIGAM™ is an IFN- γ release assay which is widely used in different national tuberculosis eradication programs world-wide. In 2015 OIE approved BOVIGAM™ for use as a primary test. Resolution 29/2014 recommends the use of BOVIGAM™ utilizing Lelystad PPD due to the improved sensitivity whereby specificity remains equivalent in comparison to PPD from CSL origin. However, test accuracy is dependent upon standardized and harmonized batch production of the applied PPD tuberculin for the stimulation of the whole blood samples.

An optimized and more standardized PPD tuberculin for IFN- γ release assay applications should be developed to improve the national tuberculosis program which is urgently needed by the cattle industry.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture (USDA), Animal and Plant Health Inspection Service (APHIS), Center for Veterinary Biologics (CVB) to work with USDA-APHIS Veterinary Services (VS) Cattle Health staff to optimize purified protein derivative tuberculin for interferon-gamma release assays and that the resulting product(s) be submitted to APHIS-VS-CVB for licensing purposes.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 43

APPROVED

SOURCE:

COMMITTEE ON SHEEP AND GOATS

SUBJECT MATTER:

Ensuring Sound Science-Based Animal Health Policies

BACKGROUND INFORMATION:

Science-based animal health policies are fundamental to the United States Department of Agriculture in their efforts to issue decisions, develop regulations, and identify diagnostic test needs.

In some cases, policies and decisions are justified by in-house studies that are not subject to a rigorous outside and independent scientific review. In other cases, potential conflicts of interest develop when a study that serves as the foundation for a regulatory decision has been published in a scientific journal with an editorial board that includes individuals from government agencies through which the decision will be issued.

Clear and sound evaluation criteria for scientific studies used to support federal animal health policies are key to retaining public trust and confidence in agency actions.

RESOLUTION:

To ensure the development of science-based animal health policy, the United States Animal Health Association urges the United States Department of Agriculture (USDA), the United States (US) Department of Homeland Security and the US Department of Interior to establish Department-wide criteria for evaluating research that is used to support animal health policy decisions. The following actions would help establish sound evaluation criteria and help increase trust and confidence in the policy making process:

- Initiate an independent and unbiased review of the science and/or methodologies used to support broad policy decisions.
- Establish a validation process for prediction models, risk assessments, spread models, or other diagnostic or analytical methods that are to be used.
- Require any studies proposed to be undertaken by an agency or department, intended to be used to justify or direct animal health policy or decision making, be subject to an independent scientific review which would be consistent with a previously established rigorous outside, independent review processes in place for evaluation of competitive grant proposals at USDA.



UNITED STATES ANIMAL HEALTH ASSOCIATION

2016 Resolution

120th Annual Meeting

October 13-19, 2016 ~ Greensboro, NC

RESOLUTION NUMBER: 44

APPROVED

SOURCE:

COMMITTEE ON PARASITIC AND VECTOR BORNE DISEASES

SUBJECT MATTER:

Development of Cattle Fever Tick Prevention and Treatment Methods for Both Livestock and Wildlife

BACKGROUND INFORMATION:

The Texas Cattle Fever Tick Eradication Program (CFTEP), established in 1906, is the oldest livestock pest eradication program in the nation. CFTEP's mission is to eradicate fever ticks through the management of a permanent quarantine zone, as well as through temporary quarantine areas created to address the presence of fever ticks outside the permanent quarantine zone. Since the onset of the Program, the required 100% treatment of cattle has been the most effective method of eradicating ticks from infested premises. The 100% treatment requirement, while primarily responsible for the successful eradication of fever ticks from the U.S. in 1946, creates a burden for producers by increasing gathering frequency and handling of cattle.

Treatment for cattle fever ticks has historically been accomplished by the application of acaricides through the use of swim vats. Multiple acaricides have been used over the years. Due to environmental concerns and tick resistance issues, coumaphos is the only remaining, licensed topical acaricide for use in eradication efforts and has a required treatment interval of 7-14 days. Doramectin is the only approved systemic acaricide and has a required treatment interval of 25-28 days. Systematic treatment of infested cattle must occur at the frequency prescribed by one of the two treatments for the duration of the quarantine period. Quarantine periods for infested cattle can last nine months or longer.

Moving forward, the key to mitigating the risk of fever tick incursions from Mexico and reducing the size of cattle fever tick outbreaks will be development and implementation of preventive therapies such as vaccines. A recently developed fever tick vaccine is now in use in beef cattle in the permanent quarantine zone and temporary preventive quarantine areas. The vaccine will be a valuable tool in eradication efforts. While it is highly efficacious against the *Rhipcephalus annulatus* tick, it has only moderate efficacy against the *R. microplus* tick, the species of fever tick involved in the current large outbreaks.

Wildlife, such as white-tailed deer, and exotic wildlife, such as red deer, elk, and nilgai antelope, are also very competent fever tick hosts. Expanding populations of these wild and exotic hosts have led to, and are continuing to be major contributors to, fever tick outbreaks outside of the permanent quarantine zone. The most recent of these includes

the current outbreak in Cameron and Willacy counties in Texas. An approximately 223,000 acre temporary preventive quarantine area was established in October 2014 in Cameron County after the discovery of infested cattle on three premises outside of the permanent quarantine zone. Since October 2014, the number of infested premises in Cameron and Willacy Counties has risen to nearly 40 and the number of quarantined acres has risen to nearly 360,000. Approximately two-thirds of the currently infested premises are attributed to infested nilgai antelope, demonstrating that the species is an important contributor to the northern movement of the cattle fever tick. Similarly, wildlife hosts are contributing to an increase in fever tick infestations in the permanent quarantine zone as land use transitions away from cattle ranching and into wildlife only operations. There is no current treatment method for nilgai antelope or other exotic wildlife hosts, and only one approved treatment for white-tailed deer.

The diminishing number and short treatment interval of approved treatments, the limited number of new treatment and prevention mechanisms for cattle, and the limited to non-existent treatment and prevention methods for wild and exotic hosts are putting fever tick eradication efforts at risk. Additionally, as the current fever tick outbreaks spread, cattle producers are being forced to assume the additional costs of increased gathering and treatment of cattle when there is no available effective mechanism to treat infested wild and exotic hosts. The only current mechanism for control of infested exotic wildlife hosts is lethal removal.

RESOLUTION:

The United States Animal Health Association urges the United States Department of Agriculture, Animal and Plant Health Inspection Service (USDA-APHIS) to collaborate with the USDA-Agricultural Research Service to prioritize research projects to:

- 1) develop, and gain approval for use of new, systemic cattle fever tick treatment products with longer treatment intervals for cattle;
- 2) develop, and gain approval for use of new cattle fever tick treatment products for wildlife, especially nilgai antelope; and,
- 3) develop, and gain approval for use of improved cattle fever tick preventive therapies, such as vaccines, for both cattle and wildlife hosts.

Further, the United States Animal Health Association urges the USDA-APHIS to prioritize resources for cattle fever tick eradication efforts through increased support of the USDA-APHIS-Veterinary Services-Cattle Fever Tick Eradication Program.